

The Status of Soybean Rust Research in China^{*}

Shan Zhihui Zhou Xin'an

(Oil Crops Research Institute of CAAS, Wuchang, 430062)

Abstract Soybean rust caused by *Phakopsora pachyrhizi* Syd was first reported in 1899 in Jilin, a province in northeast China. It became the most destructive disease for soybean production in tropical or sub tropical areas since the 1960s. Systematic research in China started since 1970s. The disease distribution, yield loss, pathogen life cycle, host range, interaction of pathogen and host, disease epidemiology, host resistance identification and screening, genetics of rust resistance and breeding, integrated management were addressed in this paper.

Key words Soybean rust; Epidemics; Resistance mechanism; Resistance resource; Physiological race

中图分类号 S565.1 文献标识码 A 文章编号 1000-9841(2006)04-0438-07

Soybean rust was first reported in 1899 in Jilin, a province in northeast China^[11]. Then it was found in other 23 provinces in China by the end of 1980s. As early as in 1914, soybean rust had already become the most serious disease in Taiwan and became a destructive disease in tropical regions after 1960s. The risk of soybean rust attracted more attention in mainland of China when it first broke out in 1975 in Henan, a province in center part of China, where soybean rust had never been reported^[34]. As a member of the International Working Group on Soybean Rust (IWGSR), more detail research in soybean rust was conducted in China since that time.

1 Pathogen distribution and host range

It is clear that the pathogens causing soybean rust are *Phakopsora pachyrhizi* and *Phakopsora meibomia*e. The pathogens, once regarded as syn-

onymous by the early investigators, were separated by Ono et al^[18]. The teliospores of *Phakopsora pachyrhizi* were thin walled, and pale colored, irregular arranged in several layers in the sorus while those from *Phakopsora meibomia*e were relatively thick walled, moderately pigmented and arranged in vertical rows of a few spore depth in the sorus. It was concluded that *Phakopsora pachyrhizi* existed both in Asia and America while *Phakopsora meibomia*e only existed in America^[47]. *Phakopsora pachyrhizi* is the only pathogen causing soybean rust in Asian countries.

Phakopsora pachyrhizi has been found in 24 provinces in China, including Taiwan, Hebei, Sichuan, Tibet, Jilin, Shanxi, Jiangxi, Fujian, Hunan, Heilongjiang, Liaoning, Hubei, Guangxi, Guangdong, Guizhou, Yunnan, Jiangsu, Zhejiang, Anhui, Shandong, Gansu, Hainan, Henan and Shanxi^[3, 9~11, 13, 14, 25, 34, 42, 43, 46]. Tan et al. confirmed that the pathogen in 19 provinces were

* 收稿日期: 2005-08-26

课题来源: 国家科技攻关项目(2004DA525B06), 湖北省“十一五”科技攻关项目

作者简介: 单志慧(1964-), 博士, 副研究员, 研究方向大豆遗传育种和植物病理学研究。

通讯作者: 周新安博士, E-mail: xazhou@public.wh.hb.cn

Phakopsora pachyrhizi while the pathogens in other 4 provinces (Hebei, Liaoning, Jilin and Heilongjiang) had not been found again. The annual area reached about 1,333,333 h m²^[35].

It has been reported that the pathogen can infect 139 species belonging to 49 genera of leguminous plants. The natural hosts included at least 30 species belonging to 17 genera, among those 21 species belonging to 12 genera distributed in China^[25, 40, 47]. Ono et al^[18] and other investigators reported that more than 100 species were infected by *Phakopsora pachyrhizi* through artificial inoculation. *Glycine max* is a preferred host in the legumes family. Other common legume hosts are *Pachyrhizi*, *Dolichos*, *Vigna*, *Crotalaria*, and *Lupinus*. It was not clear if there was any alternative host species outside legumes.

2 Biological characteristics and physiological race differentiation

Phakopsora pachyrhizi had its own asexual and sexual stage. Uredospores and teliospores were the main spore forms at these two stages respectively. Uredospores were obovoid or broadly elliptical, minutely and densely echinulate and colorless to pale brown^[40], wall thick 1~1.5 μm, pycnium was not obvious. Uredospores from China were 29.36×17.88 μm in size in average^[27]. The size of the spores varied with regions^[25]. The life span of uredospore usually is 61 days at 13~24 °C. The vigor can be kept under -140 °C for 3 months. The spores start germinating under 8~36 °C with 100% relative humidity (RH)^[34]. Uredospore germ tube growth was anti phototropic^[27]. The pH required for spore germination was at pH 2.2~10 with the best germination ratio at pH 5~6^[27].

Teliospore, a key indicator of *Phakopsora* taxonomy, was the main spore type in the sexual stage of *Phakopsora pachyrhizi*. Telia were 2~7 spores layered, often mixed with uredinia. The size varied with the isolates origin. The shape of

teliospore was oblong to ellipsoid, light yellowish, single or cluster celled scattered or cluster arrangement in the leaf tissue^[27].

Teliospores and telia could be formed on many host plants such as *Glycine species*, *Cajanus cajan*, *Pachyrhizi*, *Phaseolus lunatus*, *Phaseolus vulgaris*, *Rhychos minina*, *Sesbania exaltata*, *Sesbania vesicaria* and *Vigna univulata*^[40, 41]. Teliospores stage has not been observed in tropical regions under natural conditions. Usually a low-temperature duration was required for teliospore formation^[40, 41]. The controlled condition for teliospores formation was 13~25 °C with 200 lux light, and the teliospore formation reached to the peak when daily average temperature was around 16 °C^[29, 31]. Teliospores usually had 3~6 months dormancy^[12, 19]. Teliospores germinated and developed into basidia and basidiospores by using dry wet alternative treatment^[12], no further development was observed. However, no teliospore germinating was observed when the experiment was repeated again in China (unpublished data).

The knowledge about teliospores is quite limited yet. It is not clear where and how teliospores germinate and develop into uredospores under natural conditions, and if there are any alternative hosts beside legumes. If there exist, how to find them. Tan^[31] suggested that teliospores might not germinate under nature conditions; they may not be the primary inoculum in north China. The primary inoculum were probably airborne from south China. However, if this was true, how to explain the rust outbreak in some temperate places, where there was no wind and rain records within 2~3 weeks, no spores caught, then rust disease seemed spread to the whole field in one night, where the pathogen came from (unpublished data)? And why rust is more seriously in autumn than in spring in the same area as the weather is favorable for pathogen multiplication in both seasons? Uncovering of mystery of teliospore will give us a more clear understanding of soybean rust.

It was noticed that isolates virulence varied from different geographic regions or legume hosts.

Lin divided 9 isolates from 3 places into 6 physiological races by using a tentative differential set consisting of six soybean varieties and 5 legumes species, however no marked differences in virulence among the isolates was observed on the soybean varieties. Ding^[4] classified 12 isolates into 5 races by using 11 soybean varieties. Yeh^[38] classified isolated from 5 areas into 3 races (1, 2, 3) by using 5 soybean genotypes. Tan^[29] inoculated isolates from Hubei on 8 soybean genotypes, identified 4 races: A, B, C and D. Among them, A and B were same as "1, 2" in Taiwan identified by Yeh. A was the most virulent isolate. C was different from "3", so the C and D were regarded as new races. Mo et al^[19] classified 4 races from the isolates in Guangxi by using 10 differentials; only one was the same as B race identified by Tan, indicating the complexity of pathogen population structure in different geographic regions. So far there has been no a set of standard differential for pathogen isolates virulence identification. The research on the virulence of pathogen isolates was restricted to the limited scales or regions.

3 Rust epidemic

Rust epidemic was caused by its multi cyclic foci and copious production of uredospores^[1]. Uredia formed 10 ~ 14 days after uredospores infected host plants, and then the released new uredospores infected the surrounding tissues again. In the regions of 19 ~ 27°N, uredospores from natural hosts such as *Glycine max*, *Glycine soja*^[47], *Glycine tomentosa* Benth^[10], *Pachyrhizus erosus* Urban^[47], *Pueraria lobata* (Willd) Ohwi^[47], *Pueraria tonkinensis* Gagnep^[47] could become the primary inoculums. However, the above hosts could not be grown in north China (north from 27°N) during the winter. Whether or not some perennial legumes such as *Apio carnea*, *Aphicarpaea trisperma* Bake, *Desmodium elegans* DC and *Desmodium williamsii*^[46] can become the over season hosts in north China still needs further investigation. Seed or soil transmission has been proved to be impossi-

ble for the pathogen spreading^[39].

The dispersal of fungal spores by the wind was thought to be the primary inoculums in the regions in north from 27°N in China^[31]. The evidence was obvious in hilly land. The pathogen infested center firstly formed in southern slope, then the infested area expanded to north part in ' < ' type with the wind. However, it was difficult to explain why rust occurred only in autumn or occurred seriously on autumn sown soybean or winter sown soybean in south China.

Climate played an important role in rust spreading. The daily average temperature at 15 ~ 26 °C was favor for uredospores germination. Rainfall and rainy days mainly influenced the development of rust in inland and flatlands^[27]. Rust occurrence was delayed during the less rainfall season whereas the disease outbreak was shifted to an earlier time and spread rapidly when the weather was continuous cloudy and drizzly. In coastland areas and mountain areas, fog or dew occurred frequently through year. A long duration of dew or fog weather played an important role in rust development in Fujian, Hainan and Yunnan where the disease was prevalent^[34].

Elevation also influenced the disease development. Disease severity was reported to be positive related with elevation below 600m, whereas such relation disappeared when the elevation was above 600m because rust was mostly seriously^[17].

Soybean could be infected in whole growing stage, but the flowering stage was more sensitive to the pathogen^[28]. Infested rate was positively related with plant growth stage^[34].

The disease severity in field also varied with landform, drainage condition, irrigation patterns, and plant density. The conditions that increased the humidity in the field such as lowlands, high plant density^[34], blocked drainage, and flood irrigation would enhance the disease severity.

Combined the meteorological information with rust investigation, Tan et al.^[35] divided soybean production area in China into three regions: heavy rust regions, frequent rust regions and occasional

rust regions. Most heavy rust areas covered the regions of 19 ~ 27° N. Rust occurred every year and resulted in yield loss except drought years, the yield loss in normal year reached 30% ~ 50%, serious yield loss reached 50% ~ 70% or even 100% when the weather was favor for rust development. The frequent rust areas were distributed in the regions of 27 ~ 34° N. When the rainy days during flower stage were more than 15 days, and monthly rainfall was above 150mm, rust can result in 50% ~ 70% yield loss. In regions north from 34° N, rainfall was less, so rust just occurred occasionally.

4 Identification and screening for rust resistance

The common evaluation method in field is the 3 digital rating system that was established by IWGSR^[36]. It is easy to be manipulated for large scale screening.

Rust resistance rate was determined according to the spots type (RB, O and Tan type), amount of uredospores multiplication in host leaves^[1] in field or in lab. However, this method could identify typical resistance genotypes. Some RB type spots also produced a lot of spores when the inoculated leaves were cultured for long time especially in field. Thus, Tan et al.^[33] divided 5 response types: immune (IM), high resistant (HR), moderate resistant (MS), moderate susceptible (MS) and high susceptible (HS). Their standard has been used for germplasm resistance rating in field or lab.

Combined primary screening in field plants and re identification in lab, about 11000 germplasm lines had been screened for rust resistance^[22,30]. No immune or highly resistance lines was identified. There were 83 resistant accessions (MR) obtained, most were autumn sown soybean. There were only 9 from summer sown or spring sown soybean. It was reported that the wild germplasm materials possessed a lot of resistance genes,

so far no related research have been investigated in China.

5 Rust resistance genetics, breeding and mechanism

It had been reported that there were at least 4 locus linked to soybean rust resistance^[7,8]. The resistant genotype may carry one or two dominant genes. By means of diallel analysis of PI459025 (R), AGS181(MS), AGS129(MS) and Houzimao (HS), Tan et al.^[32] confirmed that the resistance in PI459025 was controlled by one dominant gene, while the tolerance to rust in AGS181 and AGS129 was controlled by multiple genes, and the super dominance was observed in the tolerant lines.

Rust resistant landraces, such as Zhangziwu, Yushanqingpi and Jiuyuehuang, could be directly used in production but most resistant germplasm lines could not be used directly due to their poor agronomic traits, such as long maturity, low yield, logging and dark coat^[21]. Conventional rust resistance breeding started in Taiwan in 1960s. Tainung 3, Tainung 4 and Kaoshung 3 were derived from hybrid progenies of PI200492, PI200490 and PI200451, respectively^[2]. PI203970 and PI230971 were used for the breeding of AGS181, AGS182, AGS183, AGS229, and AGS233^[24]. Tan et al. used PI459025 as a parent crossed with AGS181, and some resistance lines such as R 34 and R 50 were selected. In addition, it was observed some progenies such as You 84 87, Zhongdou 19 from non resistant crossing combination behaved moderate resistance, this may be due to the improvement of horizontal resistance through hybridization. Resistant lines also could be obtained through mutation. Zaochun No. 1 was obtained from the mutation progenies of Aijiaoza, a land race in Hubei province, treated with ⁶⁰Co, and 9854 from Zhongdou 24 by γ ray irradiation^[23]. Unfortunately, there were many examples that resistant germplasm or breeding varieties lost their resistance gradually due to the emergence of new

rices^[18, 21].

Histological study showed that the leaf trichome of resistant genotypes were short and thick in regular arrangement while those on susceptible genotypes were soft and thin in irregular arrangement^[5]. The mastoid protuberance was formed in resistant genotype 3 days after inoculation on leaf surface while no mastoid protuberance formation occurred in susceptible ones. Lignification occurred much rapidly than that in susceptible ones^[9]. Isoenzymes changed obviously in amount and types after inoculation. Such responses may efficiently prevent spores dispersing on leaf surface^[5]. The acidic amino acids in resistant genotypes increased significantly 24 hour after inoculation while the neutral or alkaline amino acids in susceptible genotypes changed greatly^[20], indicating that rust resistance might possess some special features.

6 Soybean rust management

Chemical application during the potent infected stage can effectively prevent the disease. Bayleton, Plantvax and Dithane M45 were found to be effective in controlling rust. Bayleton was the best one for 90% control efficiency and over 70 days long effect duration. Daconil and Mebenil were efficient to reduce the yield loss in autumn sown soybean area^[45]. Plantvax, Benomyl, Mancozeb, Sapro (Triforine) and Triazone were also effective to control rust^[44].

Integrated protect management (IPM) includes growing resistant varieties, changing cultivation patterns such as spring sown soybean instead of autumn sown, applying potassium, decreasing the field humidity by reducing planting population, and assisted with chemical applying at early stage. The IPM strategies exert an efficient effect in soybean production in rust infected area.

7 The prospect of rust research

As has been reported in 39 countries distrib-

ted in Asia, America, Oceania, Europe and Africa, soybean rust has become a worldwide disease in soybean production. However, soybean rust possess much potential risk in production due to the complexity of the pathogens and lacking of immune or high level resistance gene source compared to other pests. Being an obligate fungus, *Phakopsora pachyrhizi* cannot be cultured under artificial conditions. In most cases, the mixtures of races were used to identify resistance accessions, the lacking of an overview about *Phakopsora pachyrhizi* differentiation made the releasing of resistance cultivars not synchronize with the evolution of the pathogen, thus reduced life span of resistance cultivars. The establishment of pathogen culture method will provide us an exact resistant spectrum for each resistant variety, avoid 'escape' of resistant lines due to improper evaluation way, facilitate our understanding about pathogen population evolution and interaction between host and pathogen. The knowledge about teliospore will help us to understand the gametic copulation, story in meiosis, new physiological race emergence, the whole life cycle of *Phakopsora pachyrhizi* and the primary inoculums in temperate regions. There has been no genotype immune to rust all over the world, screening new resistant germplasm lines in wild soybean perhaps will provide us some exciting result for soybean resistance breeding. The advances in plant functional genomic and comparative genomic will also provide some clue for rust resistance research.

References

- 1 Bromfield KR; Melching JS; King solver CH. Virulence and Aggreessiveness of *Phakopsora pachyrhizi* Isolates Causing Soybean Rust[J]. Phytopahtology 1980, 70: 17 - 21.
- 2 Cheng SH, Shanmugasundarm S. Implementation and achievements in legume crops[A] 1992 Pages 8 - 27 In AVRDC, Joint Assessment and Planning of Vegetable Research and Development. Proc. Of ARC /AVRDC Workshop. Shanhuah, Taiwan. 1992.
- 3 Cummins GB. Uredinales of continental China collected by SY Shoo Mycologia, 1950. 42: 779 - 797.
- 4 Ding J. Host range and physiological races of soybean rust

- pathogen (*Phakopsora pachyrhizi*) Shanhua, Taiwan: AVRDC 1983, 14p.
- 5 Fei PH, Tan YJ, Shan ZH. Histological study of soybean rust [J]. The Chinese Journal of oil crops in China. 1996, 18(1): 48 - 50.
 - 6 Fei PH, Tan YJ, Yu ZL, et al. The resistance mechanism study in soybean rust. [C] The 6th national soybean conference symposium. 1997.
 - 7 Hartwig EE, Bromfield KR. Relationship among three genes conferring specific resistance to rust in soybean[J]. Crop Sci. 1983, 23: 237 - 239.
 - 8 Hartwig EE. Identification of a fourth major gene conferring resistance to soybean rust[J]. Crop Sci. 1986, 26: 1135 - 1136.
 - 9 Hirastuka N. Materials for a rust flora of Menchoukou I Trans [J]. Sapporo Nat. Hist. Soc Trans. 1941, 16: 193.
 - 10 Hirastuka N. Uredinological Studies[M]. Tokyo: 1955, 392.
 - 11 Jaczewski A L. Fungi Rossiae exsiccata[C]. Fasc. V. 1899.
 - 12 Koch EL, Hoppe HH. Germination of the teliospores of *Phakopsora pachyrhizi*. [J] Soybean Rust Newsletter. 1987, 8: 3 - 4.
 - 13 Ling L. Host index of the parasitic fungi of Szechuan[J]. Plant Dis Rep Suppl. 1948, 173: 1 - 38.
 - 14 Luo QZ. Primary study of plant diseases in Shaowu urban[J]. Journal of Xiehe university, 1941, (3): 357 - 369.
 - 15 Michigan Department of Agriculture. Soybean rust action plan [R], Michigan Department of Agriculture, Michigan state University. 2005, 28p.
 - 16 Mo JY, Huang FN, Zhu GN, et al. The primary study in physiological races differentiation of *Phakopsora pachyrhizi* isolates in Guangxi[J]. Guangxi Agriculture Science, 1997, 5: 239 - 241.
 - 17 Mo JY, Zhu GN, Li KH, et al. Occurrence and disease development of soybean rust in Guangxi[C]. Advance of soybean rust research, Hubei Science and Technology Press. 1994a, 77 - 83.
 - 18 Ono Y, Bunica P, Hennen JF. Delimitation of *Phakopsora*, *Phytophthora* and *Cerotelium* and their species on Leguminosae[J]. Mycological Research. 1992, 96(10): 825 - 850.
 - 19 Saksirat W, Hoppe HH. Teliospore germination of soybean rust fungus (*Phakopsora pachyrhizi* Syd). wild *Glycine* spp[J]. Journal of Phytopathology. 1991, 32: 4: 339 - 342.
 - 20 Shan ZH, Fei PH, Tan YJ. The changes of amino acid between resistant and susceptible germplasm after *Phakopsora pachyrhizi* inoculation[C]. The Second Young Scientist Symposium in Hubei Province. 1996.
 - 21 Shan ZH, Tan YJ. Genetic diversity of soybean germplasm resistant to rust[C]. Proceedings of the first Asian Conference on Plant Pathology. 2000b, (8), 25 - 28, 228.
 - 22 Shan ZH, Tan YJ, Shen MZ. Evaluation of soybean germplasm for rust resistance in China[J]. The Chinese journal of oil crops. 2000a, (4): 62 - 63.
 - 23 Shan ZH, Tan YJ, Shen MZ. The mutation effect of Co⁶⁰ - γ ray on soybean seed. Hubei Plant Protection Congress. 1999.
 - 24 Shanmugansundaram S, Yan MR, Wang TC. Breeding for soybean rust resistance in Taiwan[R]. VII World Soybean Research Conference IV international soybean processing and utilization conference III congresso Mundial de Soja (Brazilian Soybean Congress) February 29 to March 5 2004. 456 - 462.
 - 25 Tai FL. Compendium of fungus in China[M], Beijing: Science press 1979.
 - 26 Tan YJ, Sun YL. Preliminary study on physiological races of *Phakopsora pachyrhizi* Syd[J]. Soybean Science. 1989b, 8(1): 71 - 74.
 - 27 Tan YJ. Epidemics and prevention of soybean rust[J]. The Oil Crops in China. 1982, 4: 1 - 8.
 - 28 Tan YJ. Epidemiology of soybean rust in China. In: Napomphet B Subhadradandhu S. (ed.) [C] New frontiers in breeding research: proceedings of the fifth international congress society for the advance of breeding researches in Asia and Oceania. Bangkok: Kasetsart University. 1986, 813 - 822.
 - 29 Tan YJ, Fei PH, Shan ZH. The formation of teliospore of (*Phakopsora pachyrhizi* Sydow) in soybean[J]. The Chinese journal of oil crops. 2001a, 23(1): 56 - 59.
 - 30 Tan YJ, Shan ZH, Shen MZ, et al. Evaluation of soybean germplasm of China for resistance to soybean rust[J]. Soybean Science. 1997, 16(3): 205 - 209.
 - 31 Tan YJ, Shan ZH, Zhou LC. The role of teliospores of *Phakopsora pachyrhizi* Sydow in soybean rust infection cycle[J]. The Chinese Journal of Oil Crops. 2001b, 23(3): 49 - 51.
 - 32 Tan YJ, Sun YL, Shan ZH. Soybean rust resistance inheritance. The Oil Crops in China[J]. 1991, 13(2): 104 - 109.
 - 33 Tan YJ, Sun YL, Yu ZL, et al. Soybean varietal reaction to rust caused by *Phakopsora pachyrhizi* Syd[C]. Advance of Soybean Rust Research, Hubei Science and Technology, 1994b, 145 - 148.
 - 34 Tan YJ, Yu ZL, Liu JL. Report of soybean rust investigation and chemicals protection[J]. The Oil Crops Research Institute of CAAS. 1976 - 1980, V16.
 - 35 Tan YJ, Yu ZL, Sun YL. Distribution and damage of soybean rust in China. Advance of soybean rust research[C]. Hubei Science and Technology Press, 1994a, p29 - 35.
 - 36 Yang CY. The IWGSR rust rating system[J]. Soybean Rust Newsletter, 1977, 1(1): 4 - 6.
 - 37 Yang CY. Soybean rust caused by *Phakopsora pachyrhizi* Presented at the first soybean rust workshop held March 21 - 27 1991 in Wuhan, Hubei China 1991, 29pp.
 - 38 Yeh CC. Differential reactions of *Phakopsora pachyrhizi* on soybean in Taiwan IN: Soybean in tropical and subtropical cropping systems[C]. Proceedings of Shanhua, Tainan, 1985, AVRDC Cp247 - 250.
 - 39 Yeh CC, Sinclair JB, Tschanz AT. *Phakopsora pachyrhizi* not transmitted by infested soybean seeds or soil[J]. Soybean Rust Newsletter, 1982a, 5(1): 44 - 47.
 - 40 Yeh CC, Tschanz AT, Sinclair JB. Induce teliospore formation by *Phakopsora pachyrhizi* (soybean rust) ten hosts[J]. Phytopathology 1981, 71(8): 914.
 - 41 Yeh CC, Tschanz AT, Sinclair JB. *Phakopsora pachyrhizi*:

- uredinal development, uredospore production and factors affecting teliospores formation on soybeans[J]. Australian Journal of Agricultural Research 1982b, 33(1): 25-31.
- 42 Yu ZL, Tan YJ, Wang YL. Soybean rust investigation report in Yunnan, Guizhou and Sichuan[J]. The Oil Crops Research Institute of CAAS. 1990.
- 43 Yu ZL, Sun YL. Soybean rust investigation in Shanxi and Gansu. The Oil Crops Research Institute of CAAS. 1991b.
- 44 Zhang XX, Zhang XN, Lin SS, et al. Study of occurrence and control of soybean rust in Jiangxi[J]. Jiangxi Agriculture Science 2004, 12(3): 41-44.
- 45 Zhang ZM, Lin ZQ, Fang QF. Preventive effect of some chemicals on the rust of autumn sown soybean[C]. Advance of soybean rust research, Hubei Science and Technology Press 1994, 123-130.
- 46 Zhuang JY. Uredinals from East Himalaya[J]. Acta Mycol. Sin. 1986, 5: 75-85.
- 47 Zhuang JY. Soybean rust in China: Pathogen, Hosts and Distribution[J]. The Oil Crops in China 1992, 3: 67-69.

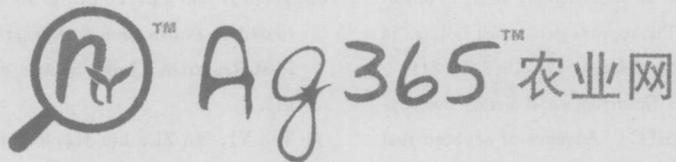
中国大豆锈病研究

单志慧 周新安

(中国农业科学院油料作物研究所, 武昌 430062)

摘要 1899年在中国吉林首次报道了由豆薯层锈菌(*Phakopsora pachyrhizi* Syd)引起的大豆锈病。20世纪60年代大豆锈病成为热带、亚热带地区大豆生产中最严重的病害。中国于20世纪70年代开始了较为系统的研究,本篇综述介绍了中国在大豆锈病的病原菌分布及其寄主、产量损失,病害流行、病原生活周期、病原与寄主的互作、抗锈资源鉴定、抗锈遗传、抗锈育种和综合防治等方面的研究进展。

关键词 大豆锈病; 流行; 抗性机制; 抗锈资源; 生理小种



网址大全

供应求购

农业资讯

企业库

养殖技术

种植技术

农业搜索



Ag365农业网(Ag365.com)是在中国农业网址大全(ny3721.com)、中国农业商务网(sw.ag365.com)等八大知名农业网站基础上整合和改版而成的农业搜索门户。

Ag365以搜索技术为核心,为农业用户提供最简单、最方便、最实用的信息查询途径,为农业企业提供最佳的产品展示平台。

Ag365正在招募授权代理商

地址: 北京市海淀区西直门北大街41号天兆5B#302 电话: 010-62273253 传真: 010-62250804-602

邮编: 100044 网站: <http://www.Ag365.com> 电子邮箱: support@Ag365.com